## Notes

## Synthesis of Diaminosuberic Acid **Derivatives via Ring-Closing Alkyne** Metathesis

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## Introduction

The side chains of cysteine residues in peptides and proteins are often involved in disulfide linkages (viz. cystine (1) in Figure 1). The presence of one or more covalent disulfide bridge(s) is important for the structural and functional properties of many polypeptides. Cystine isosteres have been developed to improve the stability of biologically active peptides since disulfide bonds are chemically and metabolically labile. The most frequently exploited isostere is the dicarba analogue (2S,7S)-2,7 -diaminosuberic acid (DAS, 2)<sup>1</sup> which has been synthesized by several groups.<sup>2–6</sup> A straightforward approach to DAS proceeds via ring-closing alkene metathesis (RCM). As shown independently by three groups,<sup>6</sup> RCM of two suitably tethered 2-amino-4-pentenoic acid derivatives led to mixtures of the corresponding (E)- and (Z)cycloalkenes, which were conveniently transformed into

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DAS via reduction of the double bond. Not surprisingly, the higher conformational freedom of the alkyl chain of DAS in comparison with disulfide linkages appeared an incentive for the development of conformationally restricted cystine isosteres.<sup>7</sup> In pursuing a similar goal, we here report the synthesis of the constrained analogues 3 and **4** (Figure 1) containing an Z-alkene or alkyne functionality, respectively.

Recently,<sup>8</sup> an efficient ring-closing alkyne metathesis (RCAM) reaction using the tungsten-alkylidyne complex (<sup>t</sup>BuO)<sub>3</sub>W≡C<sup>t</sup>Bu (5)<sup>9</sup> as the precatalyst has been introduced to produce cycloalkynes from the corresponding Me-terminated dialkynes. In the process of creating macrocyclic rings, RCAM has the advantage over RCM that the formation of geometrical isomers is circumvented. Having developed an efficient and reliable biocatalytic method for the synthesis of unsaturated amino acids in both enantiomerically pure forms, <sup>10</sup> we envisaged that application of tailor-made acetylene-containing amino acids in a tungsten-catalyzed RCAM strategy would give access to cystine isosteres 3 and 4.

Enantiopure (S)-2-tert-butoxycarbonylamino-hex-4ynoic acid (*S*)-(**6**) was condensed in a two-step procedure with either 1,2-ethanediol, 1,2-, or 1,3-benzenedimethanol to afford (Scheme 1) the corresponding tethered adducts (S,S)-10, (S,S)-11, and (S,S)-12, in satisfactory overall yields. RCAM of the latter individual compounds mediated by the tungsten catalyst 5 (6 mol %, chlorobenzene, 3 h, 80 °C) led to the cyclic acetylenes (*S*,*S*)-13, (*S*,*S*)-14, and (S,S)-15 in 48, 66, and 84% yield, respectively. Small amounts of cyclodimeric products were also detected. The substantial increase in yield of 15 may be attributed either to the larger ring size or to a more favorable

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|            | used in step 1   | used in step 2         | (yield)                                  | (yield)                                  |
|------------|------------------|------------------------|--|--|
| ζ          | ( <i>S</i> )-6   | ( <i>S</i> )- <b>6</b> | ( <i>S,S</i> )- <b>10</b> (84%)          | ( <i>S</i> , <i>S</i> ) <b>-13</b> (48%) |
| $\bigcirc$ | ) ( <i>S</i> )-6 | ( <i>S</i> )- <b>6</b> | ( <i>S</i> , <i>S</i> )- <b>11</b> (78%) | ( <i>S</i> , <i>S</i> )- <b>14</b> (66%) |
| 6          | ( <i>S</i> )-6   | ( <i>S</i> )- <b>6</b> | ( <i>S</i> , <i>S</i> ) <b>-12</b> (54%) | ( <i>S</i> , <i>S</i> )- <b>15</b> (84%) |
| $\langle$  | ( <i>S</i> )-6   | ( <i>R</i> )-6         | ( <i>S</i> , <i>R</i> )-10 (81%)         | ( <i>S</i> , <i>R</i> )- <b>13</b> (61%) |
| $\propto$  | ) ( <i>R</i> )-6 | ( <i>R</i> )-6         | ( <i>R</i> , <i>R</i> )-11 (78%)         | ( <i>R</i> , <i>R</i> )-14 (60%)         |
| 5          | ( <i>R</i> )-6   | ( <i>R</i> )-6         | ( <i>R</i> , <i>R</i> )-12 (68%)         | ( <i>R</i> , <i>R</i> )- <b>15</b> (80%) |
|            |                  |                        |  |  |

orientation of the alkylidene functionalities in adduct 12 in comparison to 10 and 11. To investigate if no epimerization took place during the ring closure conditions, we also prepared the meso-derivative (S,R)-10. To this end, enantiopure (S)-6 and (R)-6 were tethered with ethylene glycol following a two-step procedure as described for the synthesis of (S,S)-7. Subjection of (S,R)-10 to the tungstenmediated cyclization conditions gave (S,R)-13. Comparison of the <sup>13</sup>C NMR data of compounds (S,S)- and (S,R)-**13** showed in each case a single set of peaks indicating, within the limits of the <sup>13</sup>C spectra, that no epimerization had taken place.<sup>11</sup> Additionally, starting from (R)-2 -tertbutoxycarbonylamino-hex-4-ynoic acid, adducts (R,R)-11 and (R,R)-12 were synthesized and cyclized under the action of the tungsten catalyst 5. Thus, cyclic compounds (R,R)-14 and (R,R)-15 were obtained in a similar yield as their enantiomers.

At this stage, the cyclic alkynes were transformed (Scheme 2) into several cystine isosteres. Lindlar reduction of cycloalkyne (*S*,*S*)-**13** followed by cleavage of the ester linkages and deprotection of the Boc protecting groups with TFA provided *Z*-alkene **3**. In addition, saponification of (*S*,*S*)-**13** with LiOH followed by cleavage of the Boc groups gave the acetylenic cystine isostere **4** in good overall yield. Alternatively, as an illustration that this approach can provide access to several diastereoisomers of diaminosuberic acid derivatives, exhaustive catalytic hydrogenation of (*R*,*R*)-**14** gave **18**,<sup>2a</sup> and subsequent removal of the Boc protecting groups furnished (2*R*,*TR*)-2,7-diaminosuberic acid, the analytical data of which were in full accord with those previously reported.<sup>3b</sup>



To incorporate the diamino dicarboxylic acids **2**, **3**, or **4** into peptides, the availability of orthogonally protected derivatives is required. For instance, the orthogonally protected adduct **22** was prepared from (*R*)-2-*tert*-butoxy-carbonylamino-hex-4-ynoic acid (*R*)-**6** in four steps by a slight modification of the route reported by Williams et al.<sup>6b</sup> (Scheme 3). Thus, tungsten mediated cyclization of **22** gave cycloalkyne **23** in 66% yield, showing that all protective groups were compatible with the RCAM process. Catalytic hydrogenation of **23** and subsequent protection of the resulting amine with an Fmoc group finally furnished the orthogonally protected (2*R*,7*R*)-2,7-diaminosuberic acid derivative **24** in 76% yield.

In summary, the ring closing alkyne metathesis reaction of 2-amino-4-hexynoic acid derivatives catalyzed by **5** proved to be a useful tool for the synthesis of several cystine isosters. The methodology allows the asymmetric preparation of saturated and unsaturated cystine isosteres as the free diamino dicarboxylic acids or in orthogonally protected form. While the availability of both enantiomers of the starting amino acids ensures access to all possible diastereomers, selective reduction of the triple bond allows the synthesis of geometrically pure conformationally restricted DAS derivatives. Adaptation of this approach to the synthesis of orthogonally protected derivatives of **3** and **4** will be reported in due course.

## **Experimental Section**

**General Methods.** Unless otherwise indicated, all reagents were obtained from commercial suppliers and were used without further purification. Solvents were dried according to established procedures by distillation from an appropriate drying agent under an inert atmosphere. Reactions involving air- or moisturesensitive reagents or intermediates were performed under argon in glassware that had been flame-dried. Flash chromatography was performed using Merck silica gel 60 (230–400 mesh). ESI mass spectra were collected by constant infusion of the sample dissolved in methanol/water with 1% HOAc. ESI is a soft ionization technique, resulting in protonated, sodiated species in positive ionization mode and deprotonated, chlorated or [M + acetic acid]<sup>-</sup> in negative ionization mode.

**General Procedure I:** Synthesis of Monoesters. *N*,*N*-Dicyclohexylcarbodiimide (1.3 equiv) was added to a solution of (*S*)-**6** (or ( $\vec{R}$ )-**6** when appropriate), the diol tether (3–5 equiv), and DMAP (0.14 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (0.04 M) at 0 °C. The reaction mixture was allowed to warm slowly and stirred overnight at room temperature. The solid was filtered off, and the solvent was concentrated under reduced pressure. EtOAc was added to

<sup>(11)</sup> A <sup>13</sup>C NMR spectrum of a 1:1 mixture of (S,S)- and (S,R)-13 was measured in CDCl<sub>3</sub> and compared with the <sup>13</sup>C NMR spectra of the reaction products (S,S)- and (S,R)-13. While the <sup>13</sup>C NMR spectrum of the 1:1 mixture showed a double set of peaks, only a single set of peaks was observed in the <sup>13</sup>C NMR spectra of (S,S)- and (S,R)-13, indicating (within the detection limit of the <sup>13</sup>C NMR ( $\leq 5\%$ )) that no epimerization took place under the reaction conditions. The <sup>13</sup>C NMR spectra of (S,S)-, (S,R)-13, and the mixture of both compounds are available as Supporting Information.



the reaction mixture, the solid was filtered off again, and the solvent concentrated. The crude product was purified by column chromatography ( $20\% \rightarrow 100\%$  ether/petroleum ether).

**General Procedure II: Synthesis of Diesters.** A solution of DMAP (3 equiv), DMAP·HCl (2 equiv), and *N*,*N*-dicyclohexy-lcarbodiimide (2 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL/mmol DMAP) was refluxed for 10 min. A solution of (*S*)-**6** (1 equiv) (or (*R*)-**6** when appropriate) and the monoester compound (obtained following the general procedure **I**) in CH<sub>2</sub>Cl<sub>2</sub> (16 mL/mmol compound) was added to the reaction mixture and stirred under reflux for 16 h. The reaction mixture was washed with 1 M HCl/H<sub>2</sub>O (1:5) and saturated aqueous NaHCO<sub>3</sub>/H<sub>2</sub>O (1:5). The organic layer was dried (MgSO<sub>4</sub>) and concentrated and the crude product purified by column chromatography (20%  $\rightarrow$  100% ether/petroleum ether).

General Procedure III: Ring-Closing Alkyne Metathesis (RCAM) Reaction. The reaction was performed under an argon atmosphere. To a solution of 5 (6% mol) in dry  $C_6H_5Cl$  (0.02 M final concentration) was added the diyne dissolved in  $C_6H_5Cl$ , and the reaction mixture was stirred at 80 °C for 1-3 h. The solvent was then concentrated, and the crude product was purified by column chromatography (20%  $\rightarrow$  100% ether/ petroleum ether).

*rac*-2-Amino-4-hexynoic Acid Amide. A solution of diphenylketimine<sup>12</sup> (29.5 g, 0.125 mol) in THF (500 mL) was treated with NaH (5.00 g, 0.125 mol) and LiI (0.53 g, 0.013 mol). After stirring for 20 min, a solution of 1-bromo-2-butyne (20.0 g, 0.15 mol) in THF (100 mL) was slowly added over a period of 1 h. After refluxing for 48 h, the mixture was cooled and evaporated, and the residue was dissolved in saturated aqueous NH<sub>4</sub>Cl (400 mL). This solution was extracted with ether ( $3 \times 400$  mL), and the combined organic layers were dried (MgSO<sub>4</sub>) and evaporated to give the crude product (45.3 g) as a white solid, which was directly subjected to the next reaction.  $R_f = 0.63$  (50% ether in petroleum ether). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.72 (t, J = 2.5 Hz, 3H), 2.83–2.66 (m, 2H), 3.73 (s, 3H), 4.25 (dd, J = 5.2, 8.2 Hz, 1H), 7.22–7.63 (m, 8H), 7.65 (d, J = 7.2 Hz, 2H).

A solution of the crude product in ether (500 mL) was treated carefully with a 1 M aqueous solution of HCl (125 mL, 0.125 mol) at 0 °C. After vigorously stirring for 16 h, the layers were separated, and the ether layer was evaporated to give the HCl salt of the crude amino methyl ester. This was immediately dissolved in H<sub>2</sub>O (200 mL), NH<sub>4</sub>OH (300 mL of a 25% solution in H<sub>2</sub>O) was added, and the reaction mixture was stirred for 3 h. The mixture was evaporated and the residue dissolved in H<sub>2</sub>O (400 mL). The pH was raised to 9 by adding 1 M aqueous NaOH, benzaldehyde (9.77 mL, 0.095 mol) was added, and the reaction mixture was vigorously stirred for 4 h. The resulting Schiff base was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 300 mL), and the combined

organic layers were dried (MgSO<sub>4</sub>) and evaporated. The residue was dissolved in acetone (700 mL), concentrated HCl (7.8 mL, 0.2 mol) was added, and the reaction mixture was stirred for 3 h. The resulting solid was filtered off to give **2-amino-4-hexynoic acid amide** as a white powder (13.5 g, 0.083 mol, 66% overall yield). Decomp (HCl salt) >243 °C, <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  1.79 (t, J = 2.4 Hz, 3H), 2.53–2.55 (m, 2H), 3.55 (t, J = 5.8 Hz, 1H). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O)  $\delta$  5.4, 24.3, 54.4, 73.7, 85.3, 173.5. HRMS (EI) calculated for C<sub>6</sub>H<sub>11</sub>N<sub>2</sub>O 126.0793, found 126.0790.

Enzymatic Resolution of 2-Amino-4-hexynoic Acid Amide. A solution of the HCl salt of the racemic amide (12 g, 0.073 mol) in distilled H<sub>2</sub>O (80 mL) was brought to pH 9.2 with KOH, followed by addition of an 80 mM solution of MnSO<sub>4</sub> until a 1 mM concentration was reached. The H<sub>2</sub>O was added until a volume of 120 mL was reached. Then whole E. coli DH5a/ *pTrpLAP* cells<sup>10</sup> (48 mg) in a HEPES buffer (1 mL) were added. The reaction mixture was shaken at 40 °C for 24 h. The reaction mixture was brought to pH 6 by carefully adding H<sub>2</sub>SO<sub>4</sub>. The enzyme was filtered off the solution, and by adding NaOH the pH was brought at 8-9; then benzaldehyde (4.09 mL, 0.036 mol) was added, and the reaction mixture was stirred vigorously for 2 h. The resulting suspension was extracted with  $CH_2Cl_2$  (3 ×), and the combined organic layers containing the Schiff base of the (*R*)-amide were dried (MgSO<sub>4</sub>), filtrated, and concentrated. The residue was dissolved in acetone, concentrated HCl (2.8 mL, 0.036 mmol) was added dropwise, and the mixture was stirred for 2 h. The solid was filtered off to give the HCl-salt of (R)-2amino-4-hexynoic acid amide as a white solid (4.01 g, 0.024 mol, 33%): ee 99% (HPLC, Crownpak CR(+)),  $[\alpha]_D = +10.5$  (c = 1, H<sub>2</sub>O). Anal. Calcd for C<sub>6</sub>H<sub>11</sub>N<sub>2</sub>OCl: C 44.32, H 6.82, N 17.23. Found: C 44.11, H 6.76, N 17.11.

The aqueous layer containing the (*S*)-acid was lyophilized and the residue was purified over a strongly acidic (Dowex 50W) ion exchange column to give (*S*)-2-amino-4-hexynoic acid as a white solid (4.0 g, 0.032 mol, 43%): ee 99% (HPLC, Crownpak CR(+)), mp 238–240 °C.  $[\alpha]_D = -35.3$  (c = 1 in H<sub>2</sub>O). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  1.77 (t, J = 2.3 Hz, 3H), 2.76–2.77 (m, 2H), 3.83–3.85 (m, 1H). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O)  $\delta$  5.2, 23.6, 56.1, 74.7, 84.3, 175.9. Anal. Calcd for C<sub>6</sub>H<sub>9</sub>NO<sub>2</sub>: C 56.68, H 7.13, N 11.02, O 25.07. Found: C 56.33, H 6.95, N 10.77.

(*R*)-2-Amino-4-hexynoic Acid. The (*R*)-amino acid amide (2.00 g, 0.024 mol) was dissolved in a phosphate buffer (pH 8, 40 mL), lyophilized whole cells of *Rhodoccocus erythropolis* NCIB 11540 (0.4 g) were added, and the mixture was shaken at 37 °C for 3 h. The reaction was monitored with TLC. After complete conversion, the mixture was centrifuged to separate the solvent from the enzyme. The water layer was lyophilized and the residue was purified with ion exchange chromatography (Dowex 50W) to obtain (*R*)-2-amino-4-hexynoic acid (1.44 g, 0.012 mol, 92%) as a white solid: ee 99% (HPLC, Crownpak CR(+)),  $[\alpha]_D = +42.8$  (c = 1 in H<sub>2</sub>O).

(S)-2 -tert-Butoxycarbonylamino-hex-4-ynoic Acid (S)-6. (S)-2-amino-4-hexynoic acid (500 mg, 3.93 mmol) was dissolved in a mixture of water (10 mL), dioxane (20 mL) and NaOH (10 mL). Di-tert-butyl dicarbonate (1.2 equiv, 4.72 mmol) was added at 0 °C and stirring was continued at room temperature for 3 h. The dioxane was partially removed under reduced pressure, EtOAc (10 mL) was added and the water layer was acidified with KHSO<sub>4</sub> 0.1 M to pH $\approx$  4. The aqueous layer was extracted with EtOAc (4  $\times$  20 mL). The combined organic layers were dried (MgSO<sub>4</sub>) and concentrated. After purification of the crude product by column chromatography (95% ether, 5% AcOH), compound (S)-6 (705 mg, 79%) was obtained as an amorphous solid.  $R_f = 0.10$  (95% ether, 5% AcOH). mp 132–133 °C.  $[\alpha]_D =$ +3.55 (c = 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.23 (s, 9H), 1.78 (t, 3H, J = 2.5 Hz), 2.74–2.67 (m, 2H), 4.43–4.42 (m, 1H), 5.29–5.28 (m, 1H), 10.27 (s, 1H).  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 3.37, 22.65, 28.15, 52.01, 72.68, 79.35, 80.32, 155.35, 175.63. HRMS (EI): calculated for C<sub>11</sub>H<sub>17</sub>NO<sub>4</sub> 227.1158, found 227.1165.

(*R*)-2 *-tert*-Butoxycarbonylamino-hex-4-ynoic Acid (*R*)-**6.** Compound (*R*)-**6** was obtained following the same procedure as described for (*S*)-**6** starting from of (R)-2-amino-4-hexynoic acid (500 mg, 3.93 mmol) in 81% yield. All the other physical data were in accord with those reported for (*S*)-**6**.  $[\alpha]_D = -3.52$ (*c* =1, CH<sub>2</sub>Cl<sub>2</sub>).

(S)-2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid, 2-Hydroxyethyl Ester (7). Compound 7 was obtained following general procedure I starting from (*S*)-6 (100 mg, 0.44 mmol) and ethylene glycol (4 equiv, 1.76 mmol, 98  $\mu$ L). Yield 101 mg (89%).  $R_f = 0.13$  (70% ether/petroleum ether). [ $\alpha$ ]<sub>D</sub> = +15.4 (c = 1, CH<sub>2</sub>-Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.43 (s, 9H), 1.75 (t, 3H, J = 2.5 Hz), 2.58 (s, 1H), 2.62 (m, 2H), 3.79 (m, 2H), 4.24 (m, 1H), 4.36 (m, 2H), 5.33 (d, 1H, J = 6.8 Hz), <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  3.33, 22.77, 28.15, 52.45, 60.65, 66.88, 73.04, 79.30, 80.20, 155.31, 171.10. HRMS (FAB +) m/z (M + 1)<sup>+</sup>: calculated for C<sub>13</sub>H<sub>22</sub>NO<sub>5</sub> 272.1498, found 272.1496.

(*S*)-2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid, 2-Hydroxymethylbenzyl Ester (8). Compound 8 was obtained following general procedure I starting from (*S*)-6 (100 mg, 0.44 mmol) and 1,2-benzenedimethanol (3 equiv, 182 mg, 1.31 mmol). Yield 120 mg (79%).  $R_f$ = 0.35 (70% ether/petroleum ether). [ $\alpha$ ]<sub>D</sub> = -5.5 (c = 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.42 (s, 9H), 1.69 (t, 3H, J = 2.5 Hz), 2.61 (m, 2H), 4.41 (m, 1H), 4.75 (s, 2H), 5.32 (m, 3H), 7.40 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ 3.28, 22.74, 28.13, 52.35, 62.44, 64.89, 72.85, 79.34, 80.07, 127.78, 128.58, 128.78, 129.70, 133.01, 139.39, 155.18, 170.84. HRMS (FAB +) m/z (M + 1)<sup>+</sup>: calculated for C<sub>19</sub>H<sub>26</sub>NO<sub>5</sub> 348.1811, found 348.1803.

(S)-2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid, 3-Hydroxymethylbenzyl Ester (9). Compound 9 was obtained following the general procedure I starting from (*S*)-6 (100 mg, 0.44 mmol) and 1,3-benzenedimethanol (4 equiv, 244 mg, 1.76 mmol). Yield 103 mg (67%).  $R_f = 0.25$  (70% ether/petroleum ether). [ $\alpha$ ]<sub>D</sub> = -5.2 (c = 0.5, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.42 (s, 9H), 1.69 (t, 3H, J = 2.5 Hz), 2.61 (m, 2H), 4.42 (m, 1H), 4.75 (s, 2H), 5.33 (m, 3H), 7.35 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  3.34, 22.90, 28.17, 52.27, 64.76, 66.93, 72.83, 79.29, 80.02, 126.43, 126.73, 127.12, 128.61, 135.60, 141.32, 155.12, 170.89. HRMS (FAB +) m/z (M + 1)<sup>+</sup>: calculated for C<sub>19</sub>H<sub>26</sub>-NO<sub>5</sub> 348.1811, found 348.1821.

(S)- 2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid 2-((S)-2-*tert*-Butoxycarbonylamino-hex-4-ynoyloxy)ethyl Ester ((*S*,*S*)-10). Compound 10 was prepared following the general procedure **II** starting from (*S*)-**6** (1 equiv, 0.39 mmol, 88 mg) and 7 (100 mg, 0.39 mmol). Yield 177 mg (95%).  $R_f$  = 0.67 (70% ether/petroleum ether). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.42 (s, 18H), 1.72 (t, 6H, *J* = 2.4 Hz), 2.61 (m, 4H), 4.34 (m, 6H), 5.27 (m, 2H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  3.30, 22.87, 28.13, 52.12, 62.67. 72.74, 79.16, 79.91, 154.98, 170.71. HRMS (FAB +) m/z(M + 1)<sup>+</sup>: calculated for C<sub>24</sub>H<sub>37</sub>N<sub>2</sub>O<sub>8</sub> 481.2550, found 481.2540.

(S)- 2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid 2-((S)-2-*tert*-Butoxycarbonylamino-hex-4-ynoyloxymethyl)benzyl Ester ((*S*,*S*)-11). Compound 11 was prepared following the general procedure II starting from (*S*)-6 (1 equiv, 0.31 mmol, 71 mg) and 8 (108 mg, 0.31 mmol). Yield 172 mg (99%).  $R_f =$ 0.67 (70% ether/petroleum ether). [ $\alpha$ ]<sub>D</sub> = +3.5. (c = 1, CH<sub>2</sub>Cl<sub>2</sub>).  $^1\mathrm{H}$  NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.43 (s, 18H), 1.68 (s, 6H), 2.61 (m, 4H), 4.43 (m, 2H), 5.32 (m, 6H), 7.39 (m, 4H).  $^{13}\mathrm{C}$  NMR (CDCl<sub>3</sub>, 100 MHz) 3.29, 22.94, 28.15, 52.30, 64.44, 72.78, 79.26, 79.94, 128.31, 128.65, 129.54, 129.63, 133.88, 155.02, 170.66. HRMS (FAB +) m/z (M + 1)+: calculated for  $\mathrm{C_{30}H_{41}N_2O_8}$  557.2863, found 557.2858.

(S)- 2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid 3-((S)-2-*tert*-Butoxycarbonylamino-hex-4-ynoyloxymethyl)benzyl Ester ((*S*,*S*)-12). Compound 12 was prepared following the general procedure II starting from (*S*)-6 (1 equiv, 0.22 mmol, 50 mg) and 9 (76 mg, 0.22 mmol). Yield 97 mg (80%).  $R_f$  = 0.53 (70% ether/petroleum ether). [ $\alpha$ ]<sub>D</sub> = -10.1. (c = 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.44 (s, 18H), 1.71 (t, 6H, J = 2.5 Hz), 2.64 (m, 4H), 4.45 (t, 2H, J = 4.2 Hz), 5.32 (m,6H), 7.33 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) 3.34, 22.95, 28.17, 52.26, 66.69, 72.81, 79.25, 79.97, 127.69, 127.95, 128.71, 135.72, 155.06, 170.83. HRMS (FAB +) m/z (M + 1)<sup>+</sup>: calculated for C<sub>30</sub>H<sub>41</sub>N<sub>2</sub>O<sub>8</sub> 557.2863, found 557.2872.

((6.5,11.5)-11-*tert*-Butoxycarbonylamino-5,12-dioxo-1,4dioxacyclododec-8-yn-6-yl)carbamic Acid *tert*-Butyl Ester ((*S*,*S*)-13). It was prepared following the general procedure III starting from (*S*,*S*)-10 (161 mg, 0.34 mmol). Yield 68 mg (48%).  $R_f = 0.15$  (70% ether/petroleum ether).  $[\alpha]_D = +77.7$  (c = 1, CH<sub>2</sub>-Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.44 (s, 18H), 2.47 (dd, 2H, *J* = 6.2 Hz, *J* = 14.9 Hz), 2.79 (m, 2H), 4.38 (m, 2H), 4.47 (m, 2H), 4.57 (m, 2H), 5.22 (d, 2H, *J* = 7.6 Hz). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  23.55, 23.79, 28.13, 52.82, 61.96, 77.86, 80.15, 154.69, 170.72. HRMS (FAB +) m/z (M + 1)<sup>+</sup>: calculated for C<sub>20</sub>H<sub>31</sub>N<sub>2</sub>O<sub>8</sub> 427.2080, found 427.2082.

((6*R*,11*S*)-11-*tert*-Butoxycarbonylamino-5,12-dioxo-1,4dioxacyclododec-8-yn-6-yl)carbamic Acid *tert*-Butyl Ester ((*S*,*R*)-13). It was prepared following the general procedure III starting from (*S*,*R*)-10. Yield 61%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  1.45 (s, 18H), 2.50 (dd, 2H, *J* = 4.1 Hz, *J* = 14.9 Hz), 2.79 (dd, 2H, *J* = 3.1 Hz, *J* = 14.9 Hz), 4.32 (m, 2H), 4.49 (bs, 2H), 4.62 (m, 2H), 5.34 (d, 2H, *J* = 7.6 Hz). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ 23.58, 23.83, 28.17, 52.92, 62.35, 77.90, 80.15, 154.75, 170.62.

((8.5,13.5)-13-*tert*-Butoxycarbonylamino-7,14-dioxo-1,6dioxa-[1.6.4]benzocyclotetradec-10-yn-8-yl)carbamic Acid *tert*-Butyl Ester ((*S*,*S*)-14). Compound 14 was prepared following the general procedure III starting from (*S*,*S*)-11 (151 mg, 0.27 mmol). Yield 87 mg (66%).  $R_f$  = 0.46 (70% ether/petroleum ether). [α]<sub>D</sub> = +9.0. (*c* = 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 1.39 (s, 18H), 2.48 (dd, 2H, *J* = 4.9 Hz, *J* = 15.4 Hz), 2.79 (m, 2H), 4.45 (bs, 2H), 5.23 (m, 6H), 7.38 (s, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ 22.98, 28.12, 52.69, 65.50, 77.25, 80.08, 129.29, 131.64, 133.82, 155.50, 170.72. HRMS (FAB +) *m*/*z* (M + 1)<sup>+</sup>: calculated for C<sub>26</sub>H<sub>35</sub>N<sub>2</sub>O<sub>8</sub> 503.2393, found 503.2390.

((5.5,10.5)-10-*tert*-Butoxycarbonylamino-4,11-dioxo-3,12dioxa-bicyclo[12.3.1]octadeca-1(18),14,16-trien-7-yn-5-yl)carbamic Acid *tert*-Butyl Ester ((*S*,*S*)-15). Compound 15 was prepared following the general procedure III starting from (*S*,*S*)-12 (95 mg, 0.17 mmol). Yield 70 mg (84%).  $R_f$ = 0.48 (70% ether/ petroleum ether). [ $\alpha$ ]<sub>D</sub> = -71.2. (c = 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  1.45 (s, 18H), 2.68 (m, 2H), 2.80 (m, 2H), 4.49 (m, 2H), 4.82 (d, 2H, J = 12.6 Hz), 5.34 (d, 2H, J = 8.3 Hz), 5.69 (d, 2H, J = 12.6 Hz), 7.28 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ 22.89, 28.18, 51.85, 66.23, 77.82, 80.09, 125.55, 127.63, 128.27, 136.68, 155.03, 170.28. HRMS (FAB +) m/z (M + 1)+: calculated for C<sub>26</sub>H<sub>35</sub>N<sub>2</sub>O<sub>8</sub> 503.2393, found 503.2361.

(Z)-(2S,7S)-2,7-Diamino-oct-4-enedioic Acid TFA Salt (3). (S,S)-13 (72 mg, 0.17 mmol) was dissolved in EtOAc:MeOH (6: 1, 3.5 mL). Quinoline (1 equiv, 0.17 mmol, 20  $\mu$ L) was added followed by the Lindlar catalyst (29 mg). The reaction mixture was stirred under H<sub>2</sub> atmosphere at room temperature. After 48 h, an additional amount of Lindlar catalyst (29 mg) and quinoline (1 equiv, 0.17 mmol, 20  $\mu L)$  were added, and stirring was continued for 48 h under H<sub>2</sub>. Then, the mixture was filtered over Hyflo. The filtrate was washed with HCl 0.25 M (3  $\times$  10 mL) and water, dried (MgSO<sub>4</sub>) and concentrated. The crude product was suspended in MeOH (9.5 mL), LiOH (0.1 M in water, 2.2 equiv, 0.37 mmol, 3.7 mL) was added, and the mixture stirred at room temperature for 48 h. After this time, the reaction mixture was covered with a layer of EtOAc and acidified to pH  $\approx$  4 with KHSO<sub>4</sub> 0.1 M. The aqueous phase was extracted with EtOAc (3  $\times$  10 mL). The combined organics were dried (MgSO<sub>4</sub>) and concentrated. The residue was purified by column chromatography (petroleum ether–EtOAc 2:1 to EtOAc–MeOH 5:1) to give **16** (41 mg, 56%). <sup>1</sup>H NMR (D<sub>2</sub>O, 200 MHz)  $\delta$  1.42 (s, 18H), 2.43 (m, 4H), 3.96 (m, 1H), 5.51 (m, 1H).<sup>13</sup>C NMR (CD<sub>3</sub>OD, 50 MHz)  $\delta$  28.43, 30.68, 80.29, 128.39, 157.59, 175.54. To a solution of **16** (9 mg, 0.022 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (0.25 mL) was added TFA (0.25 mL), and the reaction mixture was stirred at room temperature for 10 min. Then, it was concentrated to dryness and coevaporated with toluene (2 × 5 mL) to give (*S*,*S*)-**3** (9 mg, 94%). [ $\alpha$ ]<sub>D</sub> = +18.0. (*c* = 0.2, H<sub>2</sub>O). <sup>1</sup>H NMR (D<sub>2</sub>O, 400 MHz)  $\delta$  2.71 (t, 4H, *J* = 5.8 Hz), 3.95 (t, 2H, *J* = 5.8 Hz), 5.66 (t, 2H, *J* = 5.1 Hz). <sup>13</sup>C NMR (D<sub>2</sub>O, 50 MHz)  $\delta$  28.80, 54.09, 127.87, 173.63. MS (ESI) *m*/*z* = 263 (M+ 2 H<sup>+</sup> + Ac<sup>-</sup>)

(2S,7S)-2,7-Diamino-oct-4-ynedioic Acid TFA Salt (4). Compound (S,S)-13 (90 mg, 0.21 mmol) was suspended in MeOH-H<sub>2</sub>O (1:1 v/v, 14 mL), LiOH (0.1 M in water, 2.1 equiv, 0.44 mmol, 4.43 mL) was added, and the mixture was stirred at room temperature for 72 h. After this time, the reaction mixture was covered with a layer of EtOAc and acidified to  $pH\approx 4$  with KHSO<sub>4</sub> 0.1 M. The aqueous phase was extracted with EtOAc (3  $\times$  10 mL). The combined organics were dried (MgSO<sub>4</sub>) and concentrated. The crude was purified by column chromatography (petroleum ether-EtOAc 2:1 to EtOAc-MeOH 10:1) to give compound 17 (73 mg, 87%). <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz)  $\delta$  1.42 (s, 9H), 1.46 (s, 9H), 2.45-2.48 m, 4H), 4.22-4.17 (m, 2H). <sup>13</sup>C NMR (CD<sub>3</sub>OD, 50 MHz) & 24.49, 28.53, 55.45, 79.29, 80.02, 156.89. Compound 17 (41 mg, 0.1 mmol) was treated with a mixture TFA-CH<sub>2</sub>Cl<sub>2</sub> (1:1 v/v, 1 mL) at room temperature for 10 min. Then, the reaction mixture was concentrated and coevaporated with toluene  $(2 \times 5 \text{ mL})$  to give (S,S)-4 (42 mg, 98%).  $[\alpha]_D = -3.8$ . (c = 1.2, H<sub>2</sub>O). <sup>1</sup>H NMR (D<sub>2</sub>O, 400 MHz)  $\delta$ 2.85 (m, 4H), 3.93 (t, 2H, J = 4.2 Hz).<sup>13</sup>C NMR (D<sub>2</sub>O, 50 MHz)  $\delta$  21.06, 52.36, 78.25, 171.58. MS (ESI) m/z = 261 (M + 2H<sup>+</sup> +  $Ac^{-}$ 

(2R,7R)-Diaminosuberic Acid (R,R-2). To a solution of (R,R)-14 (83.0 mg, 0.17 mmol) in EtOAc (4.0 mL) Pd/C (8.0 mg) was added. After stirring under an atmosphere of H<sub>2</sub> for 4 h, the reaction mixture was flushed with N<sub>2</sub> and filtered over Hyflo. Purification of the crude product by flash chromatography (95% ether, 5% AcOH) furnished **18**<sup>2a</sup> (68.0 mg, 99%).  $R_f = 0.05$  (70% ether/petroleum ether). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  1.50-1.30 (m, 22H), 1.70-1.50 (m, 2H), 1.85-1.70 (m, 2H), 4.05-4.04 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) & 26.50, 28.75, 32.66, 54.86, 80.48, 158.18, 176.35. A solution of 18 (68.0 mg, 0.17 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2.0 mL) and TFA (2.0 mL) was stirred for 16 h at ambient temperature. After evaporation of the solvents, the crude product was purified with ion exchange chromatography (dowex 50  $\times$  4W eluted 2 M NH<sub>4</sub>OH) which afforded (*R*,*R*)-2 (15 mg, 44%).  $[\alpha]_D = -26.8$  (c = 1, 2 M HCl) [lit. for (2S,7S)-2,7diaminosuberic acid  $[\alpha]_D = +24.8$  (c = 0.25,  $H_2O$ )<sup>3b</sup>]. <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O, HCl) & 1.53-1.45 (m, 2H), 2.00-1.91 (m, 2H), 4.05 (t, 1H, J = 6.3 Hz). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O, HCl)  $\delta$  26.54, 32.11, 55.57, 174.88.

(*R*)-2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid 3-Hydroxymethylbenzyl Ester (19). Compound 19 was prepared following the general procedure I starting from (*R*)-6 (885 mg, 3.9 mmol) and 1,3-benzenedimethanol (5 equiv, 19.5 mmol, 2.7 g). Yield 1.05 g (77%).  $R_f = 0.26$  (70% ether/petroleum ether).  $[\alpha]_D = +5.6$  (c = 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  1.32 (s, 9H), 1.59 (s, 3H), 2.52 (m, 2H), 3.68 (bs, 1H), 4.30 (m, 1H), 4.51 (s, 2H), 5.04 (m, 2H), 5.44 (d, 1H, J = 8.7 Hz) 7.18 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$  3.31, 22.81, 28.11, 52.25, 64.50, 66.93, 72.81, 79.24, 79.99, 126.39, 126.66, 126.99, 128.51, 135.45, 141.43, 155.10, 170.84. HRMS (FAB +) m/z (M + 1)<sup>+</sup>: calculated for C<sub>19</sub>H<sub>26</sub>NO<sub>5</sub> 348.1811, found 348.1841.

(*R*)-2-*tert*-Butoxycarbonylamino-hex-4-ynoic Acid 3-(4-Nitro-phenoxycarbonyloxymethyl)benzyl Ester (20). To a solution of **19** (134 mg, 0.38 mmol) in DMF (5 mL), bis(4nitrophenyl) carbonate (2.5 equiv, 0.96 mmol, 292 mg), and DIPEA (1.5 equiv, 0.57 mmol, 100  $\mu$ L) were added, and the reaction mixture was stirred for 16 h at room temperature. Then, it was poured over EtOAc (50 mL), washed with KOH 0.01 N (7 × 25 mL) and brine (25 mL), dried (MgSO<sub>4</sub>), and concentrated. The residue was purified by column chromatography petroleum ether–EtOAc 5:1 to give **20** (177 mg, 91%).  $R_f$ = 0.80 (70% ether/ petroleum ether). [ $\alpha$ ]<sub>D</sub> = +7.5. (*c* = 1, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  1.37 (s, 9H), 1.61 (s, 3H), 2.61 (m, 2H), 4.38 (m, 1H), 5.22 (m, 4H), 5.39 (d, 1H, *J* = 8.7 Hz), 7.29–8.19 (m, 8H).  $^{13}\mathrm{C}$  NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$  3.19, 20.72, 22.75, 28.03, 52.29, 66.45, 70.33, 72.88, 79.16, 79.82, 114.79, 115.36, 121.58, 125.07, 125.86, 128.01, 128.28, 128.49, 128.83, 134.53, 136.05, 145.17, 152.21, 155.06, 155.30, 170.77. HRMS (FAB +) m/z (M + 1)+: calculated for  $C_{26}H_{29}N_2O_9$  513.1873, found 513.1877.

(R)-2-[3-((R)-2-tert-Butoxycarbonylamino-hex-4-ynoyloxymethyl)benzyloxycarbonylamino]hex-4-ynoic Acid (21). To a solution of 20 (1.3 equiv, 2.25 mmol 1.15 g) in dry DMF (9 mL) were added of (R)-2-amino-4-hexynoic acid (1.7 mmol, 215 mg) and DIPEA (2.6 equiv, 4.5 mmol, 780  $\mu$ L), and the reaction mixture was stirred at room temperature for 48 h. Then, it was diluted with EtOAc (50 mL) and acidified with KHSO<sub>4</sub> 0.1 M until pH $\approx$ 4. The aqueous phase was extracted with EtOAc (4  $\times$ 50 mL). The combined organics were dried (MgSO<sub>4</sub>) and concentrated to dryness. The residue was purified by column chromatography (petroleum ether-EtOAc 4:1 to EtOAc-MeOH 10:1) to give compound 21 (820 mg, 96%). Unreacted carbonate **20** was also recovered (178 mg).  $R_f = 0.19$  (petroleum ether/ EtOAc 2:1 2% AcOH).  $[\alpha]_D = +6.2$ . (c = 0.8,  $CH_2Cl_2$ ). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz) & 1.39 (s, 9H), 1.67 (s, 6H), 2.66 (m, 4H), 4.41 (m, 2H), 5.08 (m, 4H), 5.40 (d, 1H, J = 8.7 Hz), 5.74 (d, 1H), 7.27 (s, 4H), 9.29 (bs, 1H).  $^{13}$ C NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$  3.25, 22.54, 22.85, 28.06, 52.23, 52.59, 66.39, 72.82, 73.00, 79.00, 79.19, 80.03, 86.92, 127.37, 127.65, 128.56, 135.59, 136.62, 155.21, 155.76, 170.83, 173.41. HRMS (EI): calculated for C<sub>26</sub>H<sub>32</sub>N<sub>2</sub>O<sub>8</sub> 500.2158, found 500.2169.

(R)-2-tert-Butoxycarbonylaminohex-4-ynoic Acid 3-((R)-1-tert-butoxycarbonyl-pent-3-ynylcarbamoyloxymethyl)benzyl Ester (22). To a solution of 21 (320 mg, 0.64 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (0.33 mL) was added a solution of tert-butyl 2,2,2trichloroacetimidate (3 equiv, 1.92 mmol, 419 mg) in cyclohexane (0.66 mL) followed by BF<sub>3</sub>·Et<sub>2</sub>O (1 M in CH<sub>2</sub>Cl<sub>2</sub>, 0.16 equiv, 102  $\mu$ L), and the reaction mixture was stirred at 20 °C for 5 h. Then, more tert-butyl 2,2,2-trichloroacetimidate (2 equiv, 1.28 mmol, 280 mg) was added, and stirring was continued for 3 h. The solid was filtered off, and the solvent was concentrated. A cold mixture of  $CH_2Cl_2$ :petroleum ether (1:1 v/v, 2 mL) was added, the precipitated solid was filtered again, and the solvent was concentrated. The crude mixture was purified by column chromatography (petroleum ether-acetone 6:1) to give compound 22 (295 mg, 83%).  $R_f = 0.76$  (70% ether/petroleum ether).  $[\alpha]_D =$ +10.0.  $(c = 1, CH_2Cl_2)$ . <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  1.42 (s, 9H), 1.48 (s, 9H), 1.65 (s, 6H), 2.63 (m, 4H), 4.31 (m, 1H), 4.40 (m, 1H), 5.19 (m, 4H), 5.33 (d, 1H, J = 8.7 Hz), 5.61 (d, 1H, J =8.0 Hz), 7.30 (s, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50 MHz) δ 3.31, 22.87,  $23.08,\ 27.75,\ 28.12,\ 52.25,\ 52.98,\ 66.41,\ 66.74,\ 79.90,\ 82.21,$ 127.54, 127.72, 127.81, 128.63, 135.06, 136.54, 155.43, 169.53, 170.84. HRMS (FAB +) m/z (M + 1)<sup>+</sup>: calculated for C<sub>30</sub>H<sub>41</sub>N<sub>2</sub>O<sub>8</sub> 557.2863, found 557.2830.

(6*R*,11*R*)-11-*tert*-Butoxycarbonylamino-4,12-dioxo-3,13dioxa-5-aza-bicyclo[13.3.1]nonadeca-1(18),15(19),16-trien-8-yne-6-carboxylic Acid *tert*-Butyl Ester (23). Following the general procedure for alkyne metathesis, compound 23 was obtained from 22 (217 mg, 0.39 mmol). Yield 128 mg (66%). *R<sub>f</sub>* = 0.67 (70% ether/petroleum ether).  $[\alpha]_D = +22.4$ . (*c* = 1, CH<sub>2</sub>-Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  1.45 (s, 9H), 1.47 (s, 9H), 2.44–2.94 (m, 4H), 4.25 (m, 1H), 4.45 (m, 1H), 4.83 (m, 2H), 5.55 (m, 2H), 5.84 (d, 1H), 7.33 (m, 4H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$  22.51, 27.79, 28.15, 51.44, 53.02, 65.24, 66.79, 78.13, 79.95, 82.86, 125.86, 126.25, 127.41, 128.13, 136.02, 138.14, 154.88, 155.28, 169.35, 170.75. HRMS (FAB +) *m*/*z* (M + 1)<sup>+</sup>: calculated for C<sub>26</sub>H<sub>35</sub>N<sub>2</sub>O<sub>8</sub> 503.2393, found 503.2375.

(2*R*,7*R*)-2-*tert*-Butoxycarbonylamino-7-(9*H*-fluoren-9-ylmethoxycarbonylamino)octanedioic Acid 8-*tert*-Butyl Ester (24). Compound 23 (45 mg, 0.08 mmol) was dissolved in dry MeOH (6 mL), and Pd/C (15 mg) was added. The mixture was stirred under H<sub>2</sub> atmosphere overnight at room temperature. After that, the reaction mixture was filtered over Hyflo and concentrated. The residue was dissolved in water (1 mL), and Na<sub>2</sub>CO<sub>3</sub> (2 equiv, 0.16 mmol, 17 mg) and NaHCO<sub>3</sub> (4 equiv, 0.32 mmol, 27 mg) were added. Fmoc-OSu (2 equiv, 0.16 mmol, 54 mg) dissolved in 1,4-dioxane (1 mL) was added dropwise at 0 °C. The reaction mixture was stirred at room temperature for 3 h. Then, it was diluted with EtOAc (10 mL) and acidified with KHSO<sub>4</sub> 0.1 M until pH 4, and the aqueous phase was extracted with EtOAc (3 × 10 mL). The combined organics were dried (MgSO<sub>4</sub>) and concentrated. The residue was purified by column chromatography (petroleum ether–EtOAc 3:1 → EtOAc–MeOH 10:1) to give compound **24** (35 mg, 76%). [ $\alpha$ ]<sub>D</sub> = -2.0. (c = 0.6, CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  1.42 (s, 9H), 1.79 (s, 9H), 1.63 (m, 2H), 1.79 (m, 2H), 2.63 (m, 4H), 4.22 (m, 3H), 4.42 (m, 2H), 5.26 (bs, 1H), 5.49 (bs, 1H), 7.29–7.79 (m, 8H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50 MHz)  $\delta$  24.78, 27.85, 28.24, 29.57, 32.43, 46.98, 54.23, 55.14, 66.87, 79.76, 119.78, 125.06, 126.91, 127.51, 141.10, 143.71, 155.95, 171.66. HRMS (EI m/z (M + Na)<sup>+</sup>: calculated for C<sub>32</sub>H<sub>42</sub>N<sub>2</sub>O<sub>8</sub>Na 605.2838, found 605.2847.

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**Supporting Information Available:** Copies of the <sup>13</sup>C spectra of compounds (*S*)-6, (*S*)-7, (*S*)-8, (*S*)-9, (*S*,*S*)-10, (*S*,*S*)-11, (*S*,*S*)-12, (*S*,*S*)-13, (*S*,*R*)-13, 1:1 mixture of (*S*,*S*)- and (*S*,*R*)-13, (*S*,*S*)-14, (*S*,*S*)-15, (*S*,*S*)-16, (*S*,*S*)-17, (*S*,*S*)-3, (*S*,*S*)-4, 19, 20, 21, 22, 23, 24. This material is available free of charge via the Internet at http://pubs.acs.org.

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